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ORIGINAL ARTICLE

Genetic architecture of survival and fitness-related traits in two populations of Atlantic salmon

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The additive genetic effects of traits can be used to predict evolutionary trajectories, such as responses to selection. Non-additive genetic and maternal environmental effects can also change evolutionary trajectories and influence phenotypes, but these effects have received less attention by researchers. We partitioned the phenotypic variance of survival and fitness-related traits into additive genetic, non-additive genetic and maternal environmental effects using a full-factorial breeding design within two allopatric populations of Atlantic salmon (*Salmo salar*). Maternal environmental effects were large at early life stages, but decreased during development, with non-additive genetic effects being most significant at later juvenile stages (alevin and fry). Non-additive genetic effects were also, on average, larger than additive genetic effects. The populations, generally, did not differ in the trait values or inferred genetic architecture of the traits. Any differences between the populations for trait values could be explained by maternal environmental effects. We discuss whether the similarities in architectures of these populations is the result of natural selection across a common juvenile environment.
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INTRODUCTION

The genetic architecture underlying phenotypic traits can be used to predict evolutionary trajectories. In particular, responses to selection are directly related to the amount of additive genetic variance (Falconer and Mackay, 1996). Non-additive genetic effects, on the other hand, have not been considered as important in part because they cannot be used to predict the response to selection (Lynch, 1994). However, there is an increasing evidence that non-additive genetic effects are key components of phenotypes (Crnokrak and Roff, 1995; Roff and Emerson, 2006). Furthermore, non-additive genetic effects are a cause of inbreeding depression (Crnokrak and Roff, 1999; Keller and Waller, 2002) and can be converted to additive genetic effects, for example, during a bottleneck, which can then provide genetic variation for natural selection to act on (Carson, 1990; also see Neff and Pitcher, 2008).

Phenotypic variance can also be explained by maternal environmental effects (Falconer and Mackay, 1996) and these effects can also affect evolutionary trajectories (Räsänen and Kruuk, 2007). For example, maternal environmental effects can have an impact on the rate and direction of change in response to natural selection and can generate rapid phenotypic changes in offspring traits as a result of the phenotypic plasticity of female traits (Mousseau and Fox, 1998; Räsänen and Kruuk, 2007). Consequently, understanding the contributions of all of maternal environmental effects, additive genetic effects and non-additive genetic effects is needed to fully understand evolutionary trajectories and the diversity of phenotypes.

Studies examining the architecture of traits have shown that the relative contributions of genetic and maternal effects can change during development and may be influenced by the correlation between the trait and fitness. Traits expressed during the early life stage tend to be influenced mainly by maternal effects, whereas traits expressed during later life stages are influenced increasingly by genetic effects (Kruuk *et al.*, 2008). Initial egg investments are often fully utilized during early development, leaving later life stage traits that are influenced by genetic effects (for example, Lindholm *et al.*, 2006; Evans *et al.*, 2010). For example, in mammals, maternal effects typically decline, whereas additive genetic effects remain constant (for example, Wilson and Réale, 2006) or increase during development (for example, Cheverud *et al.*, 1983). In addition, life-history traits, such as survival, that have strong correlations with fitness typically have larger non-additive than additive genetic effects, whereas morphological traits, such as body size, that have weaker correlations with fitness typically have larger additive than non-additive genetic effects (Crnokrak and Roff, 1995; Roff and Emerson, 2006). Independent of trait type, directional selection, or to some extent stabilizing selection, on traits can erode additive genetic effects, fixing alleles across loci and leaving only non-additive genetic effects (Willis and Orr, 1993). For example, morphological traits that are under strong directional selection in domestic species often have larger non-additive than additive genetic variances (Roff and Emerson, 2006).

In this study, we examine the phenotypic variance of survival and fitness-related traits at three early life-history developmental stages (egg, alevin and fry) in Atlantic salmon (*Salmo salar*). Atlantic salmon have declined sharply throughout their North American range over the past 2 centuries (Dunfield, 1985). Lake Ontario once supported the largest freshwater population of Atlantic salmon in the world, but was extirpated by 1900 (COSEWIC, 2010). Several candidate

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populations are now being considered for reintroduction into Lake Ontario based on ecological evaluations, including Atlantic salmon from the LaHave River, Nova Scotia and Sebago Lake, Maine (Dimond and Smitka, 2005). We used a full-factorial quantitative genetic breeding design to partition phenotypic variance in survival and fitness-related traits to maternal environmental, additive and non-additive genetic effects. The resultant data were used to examine the relative contributions of additive and non-additive genetic effects to morphological and life-history traits, as well as any shift in contributions during early life stages.

MATERIALS AND METHODS

Crosses

Adult broodstock fish from each source population were provided by the Ontario Ministry of Natural Resources (OMNR). For this study, LaHave ($n=25$) and Sebago families ($n=25$) were produced on 4 November 2010 at the OMNR Harwood Fish Culture Station, Harwood, Ontario, Canada following the methods of Pitcher and Neff (2006). Five females and five males from each population were mated in all possible combinations to produce full-factorial breeding design (Lynch and Walsh, 1998, p. 598). Subsamples of eggs ($n=7$) from each female were measured for diameter (nearest 0.01 mm) using digital callipers and mass (nearest 0.0001 g) using a digital scale. Those eggs were then frozen at $-20\,^{\circ}C$, transported to Western University and kept frozen for subsequent energy content analysis. Remaining eggs were randomly placed into sections of Heath-style incubators and then tanks after hatching at the OMNR Codrington Research Facility, Codrington, Ontario, using two to three sections for each full-sibling family based on offspring numbers (that is, to keep densities in sections equal). Details of the broodstock and rearing of the families are provided in Supplementary Appendix A.

Survival and fitness-related traits

We collected six measures of survival as direct measures of early life fitness: egg survival (fertilized egg to hatch, days 0–83, also examined as a rate over time); alevin survival (post-hatch until yolk sac absorption, days 84–138, also examined as a rate over time); fry survival (yolk sac absorption until released into the wild, days 139–192); and overall survival (fertilized egg until released into the wild). We also measured 12 traits that are known to be related to fitness in salmonids (Metcalfe and Thorpe, 1992; Berg *et al.,* 2001; Pakkasmaa *et al.,* 2001; Koskinen *et al.,* 2002): egg diameter and mass; egg contents at fertilization (relative fat, protein and energy); development time to hatch (also examined as a rate over time); body length at hatch; yolk sac volume at hatch; body length at yolk sac absorption; specific growth rate; and yolk sac conversion efficiency. Details on the methodology to estimate these parameters are provided in Supplementary Appendix A.

Statistical analysis of parental and population effects

All six survival and 12 fitness-related traits were examined for a population effect in addition to individual parental effects (dam and/or sire effects), position effects (tray and tank effects) and density effects using Akaike Information Criteria forward step-wise model selection in R 2.10.1 (available at http://www.r-project.org/). Statistical significance was set at $\alpha=0.05$, and all non-proportional data were checked visually for approximate normality using histograms before the analysis with parametric statistics (Crawley, 2005). Linear models were used for normally distributed data and binomial models were used for binary data (that is, one for alive and zero for dead and one for hatched and zero for non-hatched). Effects that did not cause a change in Akaike Information Criteria of greater than 10 were considered to be poorly supported and were removed from the model (Burnham and Anderson, 2002). Remaining effects were tested for significance using an analysis of variance of a linear model or an analysis of deviance of a binomial model. Non-significant effects, starting with non-significant interactions, were removed one at a time.

If individual parental effects were retained by the model selection process, the data were analyzed using mixed-effects models that treated individual parental effects as random intercepts and examined *population* as a fixed effect (in addition to the fixed effects of *density* if retained by the selection process). Any significant position effect if retained by the selection process was treated as a random intercept. Restricted Maximum Likelihood linear mixed-effects models were used for normally distributed data, and Laplace approximation binomial generalized linear mixed-effects models were used for binary data in the lme4 package of R. The mixed-effects model output in the lme4 package does not produce significance values for fixed effects; therefore, significance for the *population* effect was determined using a likelihood ratio test between the full model and a reduced model without *population*.

Statistical analysis of architecture

In addition to parental and population effects, we examined nine out of the 18 survival and fitness-related traits for architecture. The nine traits that were not examined were the overall survival measure, because we could not control for position effects (see Supplementary Appendix A), the five egg traits (that is, diameter, mass, relative fat, protein and energy), because data were collected from only one family for each female, and the three traits examined as a rate over time (that is, egg survival, alevin survival and development time to hatch), because standard analyses cannot incorporate the inclusion of a time variable. First, the phenotypic variance was partitioned into random effects for *dam ID* (V_D, maternal environmental and maternal additive genetic variance), *sire ID* (V_S, paternal additive genetic variance) and *dam ID* × *sire ID* ($V_{D\times S}$, non-additive genetic variance) components using a mixed-effects model. We used individual estimates of traits (for example, individual survival and length) to account for within-family variation, because means of family replicates overestimates genetic effects (see Puurtinen *et al.,* 2009; Neff *et al.* 2011). Means of family replicates were used for specific growth rate and yolk sac conversion efficiency because individual estimates were not available. Regardless of the Akaike Information Criteria criterion noted above, position effects were always included as a random effect to ensure that we did not overestimate non-additive genetic effects. Significances of the variance components were determined by likelihood ratio tests as above. The genetic and environmental variance components were calculated on the basis of (Lynch and Walsh, 1998, p. 509): $V_D=\frac{1}{4}\,V_A+V_M$; $V_S=\frac{1}{4}\,V_A$; and $V_{D\times S}=\frac{1}{4}\,V_N$. Negative variance components were set to a value of zero.

Using a similar method outlined in Neff and Fraser (2010), bootstrap 95% confidence intervals (CIs) were produced by first resampling with replacement the individuals within each replicate for each family until the original size was reproduced for the trait assessments. We resampled individuals to account for within-family variation and ensure that the genetic effects were not overestimated (see Puurtinen *et al.*, 2009). We resampled means per replicate for specific growth rate and yolk sac conversion efficiency because individual estimates were not available. Using the resampled data set, additive, non-additive and maternal environmental variance components were calculated as a percentage of the phenotypic variance. The resampling and calculations were repeated 1000 times and the 95% CI was determined for each parameter. In addition, pair-wise population comparisons for each metric were done by calculating for one population the proportion of comparisons that were either larger or smaller than the other population. The proportions served as one-tailed *P*-values testing for differences between populations.

RESULTS

Summary statistics of survival and fitness-related traits are presented in Table 1 and Supplementary Appendix B. Of the 'dead' eggs that were removed from the Heath trays, 99.7% (43 397 out of 43 536 eggs) were fertilized. There was nearly 100% offspring mortality for one Sebago female ($n=5$ families). Thus, the offspring from those families were not used in any of the analyses. Individual parental effects and position effects (in the Heath trays and tanks) had significant influences on survival and fitness-related traits for model selection (Table 2). These effects were subsequently treated as random effects in the mixed-effects models. The examination of architecture revealed that maternal and non-additive genetic effects explained most of the phenotypic variance in survival and fitness-related traits (Figure 1; Supplementary Appendix C).

Table 1 Summary of survival and fitness-related traits from two populations of Atlantic salmon (*Salmo salar*)

Trait	n	*LaHave*	n	*Sebago*
Egg traits				
Diameter (mm)	35	5.72 ± 0.34	28	5.33 ± 0.40
Mass (g)	35	0.1051 ± 0.0133	28	0.0864 ± 0.0168
Relative fat ($g\,g^{-1}$ of egg)	35	0.0031 ± 0.0077	28	0.0089 ± 0.0141
Relative protein ($g\,g^{-1}$ of egg)	35	0.3702 ± 0.0321	28	0.3780 ± 0.0387
Relative energy ($kJ\,g^{-1}$ of egg)	35	9.00 ± 0.76	28	9.42 ± 0.88
Egg survival (%)				
Over time	75	$-3.29 \times 10^{-3} \pm 2 \times 10^{-5}$	40	$-4.14 \times 10^{-3} \pm 3 \times 10^{-5}$
Days 0–83	75	69.1 ± 19.0	40	53.8 ± 19.9
Alevin survival (%)				
Over time	75	$-3.30 \times 10^{-2} \pm 6 \times 10^{-3}$	40	$-2.30 \times 10^{-2} \pm 5 \times 10^{-3}$
Days 84–138	75	84.0 ± 8.2	40	79.9 ± 8.8
Fry survival (%)				
Days 139–192	75	61.3 ± 19.5	40	58.0 ± 19.0
Overall survival (%)	25	35.7 ± 10.2	20	23.6 ± 14.1
Development time				
Over time	75	$2.42 \times 10^{-1} \pm 2 \times 10^{-3}$	40	$1.11 \times 10^{-1} \pm 1 \times 10^{-3}$
To hatch (degree-days)	75	479.8 ± 6.4	40	472.3 ± 12.1
Size traits				
Body length at hatch (mm)	750	16.3 ± 0.8	400	15.6 ± 0.8
Yolk sac volume (mm^3)	750	72 ± 17	400	64 ± 15
Body length at yolk sac absorption (mm)	750	25.8 ± 1.0	400	25.7 ± 1.2
Energy conversion				
Specific growth rate ($100 \times$ ln(mm)/degree-days)	75	0.146 ± 0.007	40	0.146 ± 0.009
Yolk sac conversion efficiency ($mm\,mm^{-3}$)	75	0.136 ± 0.016	40	0.158 ± 0.018

Presented are means ± 1 s.d., except for over time traits that are logit estimate ± 1 s.e. There were 25 LaHave families (5 females × 5 males) and 20 Sebago families (4 females × 5 males). Egg traits were based on seven eggs per female. Survival, development time to hatch and energy conversion numbers (*n*) represent the total number of replicates: three per LaHave family and two per Sebago family. Size traits were represented by 10 individuals per replicate. For example, *n* of 35 for LaHave egg traits is based on seven eggs from each of the five females and *n* of 750 for LaHave size traits is based on 10 individuals from each of the three replicates from each of the 25 families.

Survival

In both populations, dam effects were significant for egg survival, alevin survival (LaHave only), but not for fry survival (Supplementary Appendix C). Sire effects were not significant for either population, whereas dam × sire effects were significant for egg survival, but not for alevin survival and fry survival (Sebago only). For the LaHave population, maternal environmental and non-additive genetic effects were similar in their contribution to egg survival, but maternal environmental effects decreased, whereas non-additive genetic effects increased during the alevin and fry stages (Figure 1). On the other hand, for the Sebago population, non-additive genetic effects were larger than maternal environmental effects in their contribution to egg survival and both non-additive genetic and maternal environmental effects decreased during the alevin and fry stages. These patterns resulted in differences between populations in the genetic architecture (additive and non-additive genetic effects) of offspring survival. Sebago had significantly higher non-additive genetic effects for egg survival but lower non-additive genetic effects for fry survival than LaHave (randomization routine one-tailed $P = 0.001$). Differences were also observed between the populations for maternal environmental effects. LaHave had significantly higher maternal environmental effects for egg and fry survival (randomization routine one-tailed $P = 0.001$). There was no significant correlations between any of the egg content measures (relative fat, protein and energy) and offspring survival (Pearson correlations, $P > 0.05$ for all; data not shown).

Fitness-related traits

Dam effects were significant for the LaHave population for development time to hatch, body length at hatch, yolk sac volume, body length at yolk sac absorption, specific growth rate and yolk sac conversion efficiency (Supplementary Appendix C). Dam effects were significant for fewer fitness-related traits for the Sebago population: development time to hatch, yolk sac volume and body length at yolk sac absorption. Sire effects on the fitness-related traits were not significant in either population, whereas dam × sire effects were significant for the traits; the exceptions being for LaHave development time to hatch and body length at hatch (Supplementary Appendix C). In both populations, non-additive genetic effects explained more of the phenotypic variance than maternal environmental effects for development time to hatch, body length at hatch (Sebago only), yolk sac volume (Sebago only), specific growth rate and yolk sac

Table 2 Model selection and mixed-effects model results for survival and fitness-related traits in two populations of Atlantic salmon (*Salmo salar*)

Trait	*Model selection*	*Mixed-effects model population effect, P-value*
Egg traits		
Diameter	*dam ID*	0.022
Mass	*dam ID*	0.021
Relative fat	No effects	
Relative protein	No effects	
Relative energy	*dam ID*	0.140
Egg survival		
Over time	*Degree-days + dam ID + tray ID + sire ID + degree-days × dam ID + degree-days × sire ID + degree-days × tray ID*	<0.001
Days 0–83	*dam ID + tray ID + sire ID*	0.126
Alevin survival		
Over time	*Degree-days + dam ID + sire ID + tank ID + degree-days × dam ID + degree-days × tank ID + degree-days × sire ID*	<0.001
Days 84–138	*dam ID + tank ID + sire ID*	0.196
Fry survival		
Days 139–192	*dam ID + tank ID + sire ID*	0.451
Overall survival	*dam ID + sire ID*	0.104
Development time		
Over time	*Degree-days + dam ID + tray ID + sire ID + degree-days × dam ID + degree-days × tray ID + degree-days × sire ID*	<0.001
To hatch	*dam ID + tray ID + sire ID*	<0.001
Size traits		
Body length at hatch	*dam ID + sire ID*	0.022
Yolk sac volume	*dam ID + sire ID*	0.226
Body length at yolk sac absorption	*dam ID + tank ID + sire ID*	0.117
Energy conversion		
Specific growth rate	*dam ID + tank ID + sire ID*	0.372
Yolk sac conversion efficiency	*dam ID sire ID + tank ID*	<0.001

All mixed-effects models contained a fixed effect for population. Mixed-effects models also contained fixed effects for density and degree-days, and random effects for *dam ID, sire ID, tray ID* and *tank ID*, if these effects were identified during model selection.

conversion efficiency (Figure 1). On the other hand, maternal environmental effects explained more of the phenotypic variance than non-additive genetic effects for body length at hatch (LaHave only), yolk sac volume (LaHave only) and body length at yolk absorption. There were no significant differences between populations in the majority of the genetic architecture values for the fitness-related traits (randomization routine one-tailed $P > 0.05$), with exception that Sebago had significantly higher non-additive genetic effects for body length at hatch than LaHave (randomization routine one-tailed $P = 0.012$; Figure 1). In contrast, there were significant differences between the populations in maternal environmental effects. LaHave had significantly higher maternal environmental effects for body length at hatch, yolk sac volume and yolk sac conversion efficiency, but lower maternal environmental effects for body length at yolk sac absorption when compared with Sebago (randomization routine one-tailed $P < 0.05$). There was no significant correlations between any of the egg content measures (relative fat, protein and energy) and the fitness-related traits (Pearson correlations, $P > 0.05$ for all; data not shown).

Population differences in performance

The populations differed at only 8 of the 18 traits that we measured (Table 2). These traits included egg survival (rate only), alevin survival (rate only), egg diameter and mass, body length at hatch, development time to hatch (rate and degree-days) and yolk sac conversion efficiency. For example, egg survival for the Sebago population declined at a faster rate than the LaHave population (Table 1). The opposite pattern was detected for alevin survival. However, the differences were generally small between the populations for egg diameter (0.4 mm, 7% of the mean) and mass (0.02 g, 21%), body length at hatch (0.7 mm, 4%) and development time to hatch (7 degree-days, 2%). LaHave hatched at a faster rate than Sebago. The populations were not significantly different in body length at yolk sac absorption despite Sebago possessing a higher yolk sac conversion efficiency than LaHave (Table 1 and 2). Although survival rates differed between the populations, there were no significant differences between the populations in the final egg and alevin survival values (that is, days 83 and 138) (Table 2).

DISCUSSION

Our results detected maternal environmental and genetic effects that explained more than half (mean ± 1s.e.: 52 ± 6%) of the phenotypic variance in survival and various fitness-related traits. Maternal environmental effects were prominent at early life stages, decreased during development, and non-additive effects becoming most prominent at later life stages. Across the nine traits that we could examine the genetic architecture of, we found that non-additive genetic effects were more prominent than additive effects. Our results also revealed that the LaHave and Sebago populations did not differ, or had small differences, for the majority of traits and, similarly, did not differ in the genetic architecture of those traits. The few differences between the populations that did exist were mostly attributed to differences in maternal environmental effects.

Maternal environmental and genetic effects may be important in explaining the phenotypic variance of survival and fitness-related traits (Qvarnström and Price, 2001). We found significant maternal environmental effects in the nine traits examined for architecture, and those effects explained a mean of 14 ± 3% of the phenotypic variance across the traits. We also found sire effects in the traits, with additive genetic effects explaining a mean of 8 ± 2% of the phenotypic variance. Similarly, 16 other studies, examining some 60 different survival and fitness-related traits in natural populations, found that maternal environmental effects explained a mean of 26 ± 3% of the phenotypic variance in the traits and that additive genetic effects explained a bit less at a mean of 18 ± 3% (see references in Table 1 in Puurtinen *et al.*, 2009; also see Evans *et al.*, 2010). Collectively, these data suggest that maternal environmental effects may be the primary factor contributing to survival and fitness-related traits during early

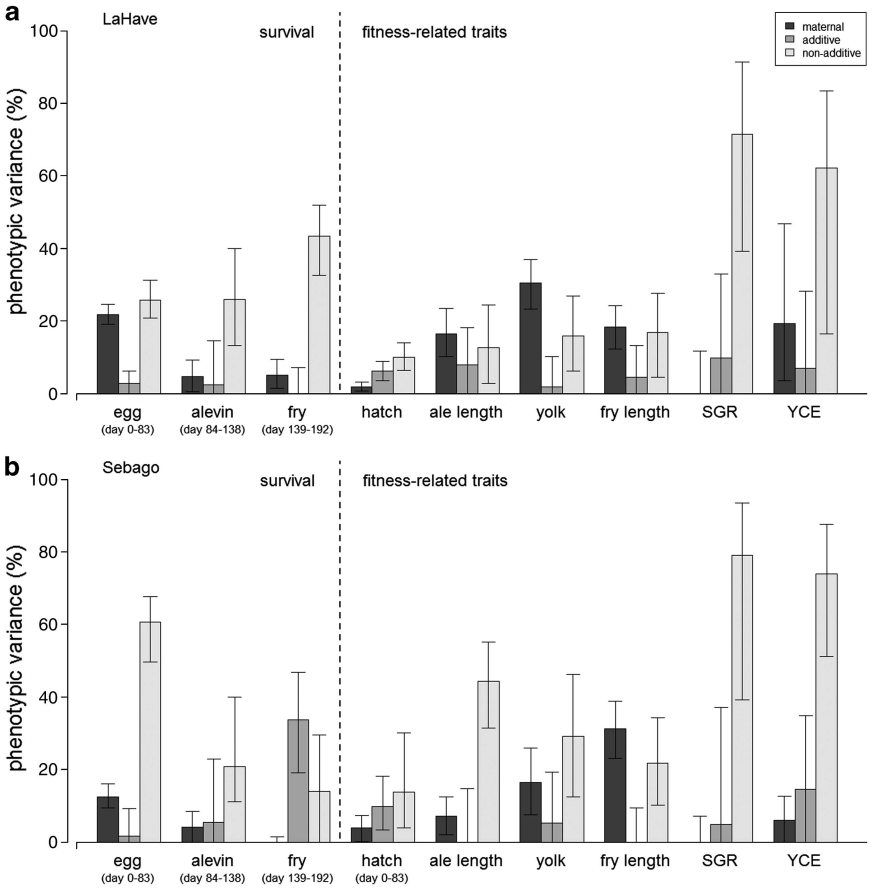

Figure 1 The architecture underlying the phenotypic variance of survival and fitness-related traits in Atlantic salmon (*Salmo salar*). Shown are data from two populations: (**a**) LaHave and (**b**) Sebago. Displayed are the median and 95% confidence intervals (CI) for maternal environmental, additive genetic and non-additive genetic effects. Hatch is development time to hatch; ale length is body length at hatch; yolk is yolk sac volume; fry length is body length at yolk sac absorption; SGR is specific growth rate; and YCE is yolk sac conversion efficiency (see Supplementary Appendix A for details).

development, although additive genetic effects also contribute to phenotypic variance during this life stage.

The amount of phenotypic variance explained by maternal environmental and genetic effects may shift during development (Fox *et al.,* 2003; Evans *et al.,* 2010). Early life stages that rely on maternal investments such as egg nutrients often have phenotypic variances explained more by maternal environmental effects (reviewed by Wilson and Réale, 2006). Later life stages instead have phenotypic variances largely explained by genetic effects, because maternal investments have been fully utilized (Wilson and Réale, 2006). We found that maternal environmental effects explained a mean of $21 \pm 3\%$ of the phenotypic variance across the four traits related to egg investments (egg survival, body length at hatch, yolk sac volume and body length at yolk sac absorption), but that genetic effects also explained a similar amount of the variance in these traits ($27 \pm 6\%$). We also found that genetic effects, largely influenced by non-additive effects, explained a mean of $47 \pm 9\%$ of the phenotypic variance across the remaining five traits that were collected after hatch. Maternal environmental effects, on the other hand, captured only $8 \pm 4\%$ of the variance in those traits. Similarly, other studies have found that maternal environmental and genetic effects explained about equal amounts of the phenotypic variance for early life stage traits (see references in Table 1 in Puurtinen *et al.,* 2009; also see Evans *et al.,* 2010). Furthermore, those studies also found that genetic effects explained $50 \pm 9\%$ and maternal environmental effects explained only $10 \pm 4\%$, on average, of the phenotypic variance for traits expressed during later life stages. Thus, the data support a shift from maternal environmental effects to genetic effects during development, but also suggest that non-additive genetic effects have an important role in survival and fitness-related traits.

Life-history and morphological traits may differ in the amount of genetic variance explained by additive and non-additive genetic effects. Life-history traits, which have strong correlations with fitness, typically have large non-additive genetic effects, whereas morphological traits, which have weak correlations with fitness, tend to have large additive genetic effects (Crnokrak and Roff, 1995; Roff and Emerson, 2006). However, a review recently suggested that additive and non-additive effects contribute about equally to both life-history and morphological traits (Puurtinen *et al.,* 2009). We found that non-additive genetic effects were larger than additive genetic effects for all nine traits that we examined for architecture, except for fry survival and development time to hatch in the Sebago population and body length at hatch in the LaHave population. Non-additive and additive genetic effects explained means of $30 \pm 6\%$ and only $8 \pm 2\%$, respectively, of the phenotypic variance across the traits. In our case, the morphological traits—body length at hatch, yolk sac volume and body length at yolk sac absorption—may have possessed larger non-additive genetic effects because these traits typically have strong

correlations with fitness in salmonids (see Koskinen *et al.*, 2002); morphological traits in other wild mammal populations typically have weak correlations with fitness (see Crnokrak and Roff, 1995; Roff and Emerson, 2006). Our data support the idea that non-additive genetic effects are larger than additive genetic effects for traits that have strong correlations with fitness and that this pattern may be independent of whether the traits are life-history or morphological in nature. Some caution is warranted when making these comparisons in our data set because our analysis is based on 5×5 crosses (albeit both populations revealed analogous patterns).

Components that explain the phenotypic variance of fitness-related traits may be preserved across populations either by shared ancestry or by parallel evolution (Schluter *et al.*, 2004). If populations possess similar traits, perhaps not surprisingly, the genetic architecture of those traits may also be similar because natural selection may have acted on the same genes that underlie the traits (Campbell and Bernatchez, 2004; Turner *et al.*, 2010; Schumer *et al.*, 2011). We found that the Atlantic salmon had similar genetic architectures (amounts of additive and non-additive effects) for seven out of the nine traits despite having a relatively large F_{ST} value. In contrast, the populations differed in the maternal environmental effects for six out of the nine traits. Similarities in the genetic architectures of these populations could be the result of parallel evolution mediated by natural selection across a common juvenile environment (that is, stream environment) (Schluter *et al.*, 2004). For example, the LaHave River and Sebago Lake stream environments have similar temperatures (J Gibson, Department of Fisheries and Oceans and M Simpson, Maine Department of Marine Resources, unpublished data), which is a key component of natural selection on fitness-related traits in the Atlantic salmon (Garcia de Leaniz *et al.*, 2007). Differences in maternal environmental effects of populations may instead reflect the differences in the adult environments (ocean vs lake) (see Räsänen and Kruuk, 2007), but more research is needed to better understand the source of those differences. Interestingly, both sources of broodstock (parents) used in our study came from a common captive hatchery environment, suggesting that the maternal environmental effects are not directly affected by the rearing environment of the dams. Regardless, the similar genetic architectures underlying the traits examined here may have resulted from parallel evolution.

In conclusion, our results detail the genetic and maternal environmental effects on phenotypic variance of survival and fitness-related traits in early life stages of two populations of Atlantic salmon. We found that the populations had similar trait values and underlying genetic architecture for the majority of traits. Our results support a shift from the maternal environmental to genetic effects during early development. They also indicate that non-additive genetic variance is as or more important for survival and fitness-related traits than additive genetic variance.

DATA ARCHIVING

Survival and fitness-related trait data have been submitted to Dryad: doi:10.5061/dryad.75030.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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Berg OK, Hendry AP, Svendsen B, Bech C, Arnekleiv JV, Lohrmann A (2001). Maternal provisioning of offspring and the use of those resources during ontogeny: variation within and between Atlantic salmon families. *Funct Ecol* **15**: 13–23.
Burnham KP, Anderson DR (2002). *Model Selection and Multimodel Inference: A Practical Information-Theoretic Approach*, 2nd edn Springer-Verlag: New York, NY, USA.
Campbell D, Bernatchez L (2004). Generic scan using AFLP markers as a means to assess the role of directional selection in the divergence of sympatric whitefish ecotypes. *Mol Biol Evol* **21**: 945–956.
Carson HL (1990). Increased genetic variance after a population bottleneck. *Trends Ecol Evol* **5**: 228–230.
Cheverud JM, Rutledge JJ, Atchley WR (1983). Quantitative genetics of development: genetic correlations among age-specific trait values. *Evolution* **37**: 895–905.
Committee on the Status of Endangered Wildlife in Canada [COSEWIC] (2010). COSEWIC assessment and status report on the Atlantic salmon *Salmo salar* (anadromous form) in Canada. *Committee in the Status of Endangered Wildlife in Canada, Ottawa, Canada*. Available at. www.sararegistry.gc.ca/status/status_e.cfm.
Crawley MJ (2005). *Statistics: An Introduction Using R*. John Wiley and Sons: Chichester, West Sussex, UK.
Crnokrak P, Roff DA (1995). Dominance variance: associations with selection and fitness. *Heredity* **75**: 530–540.
Crnokrak P, Roff DA (1999). Inbreeding depression in the wild. *Heredity* **83**: 260–270.
Dimond P, Smitka J (2005). Evaluation of selected strains of Atlantic salmon as potential candidates for the restoration of Lake Ontario. *Trout Unlimited Can Tech Rep ON-012*. Available at. www.tucanada.org/reports/ON-012_AtlanticSalmonStrains.pdf.
Dunfield RW (1985). The Atlantic salmon in the history of North America. *Can Spec Publ Fish Aquat Sci* **80**: 181 pp.
Evans ML, Neff BD, Heath DD (2010). Quantitative genetic and translocation experiments reveal genotype-by-environment effects on juvenile life-history traits in two populations of Chinook salmon (*Oncorhynchus tshawytscha*). *J Evolution Biol* **23**: 687–698.
Falconer DS, Mackay TFC (1996). *Introduction to Quantitative Genetics*. Longman: Harlow, Essex, UK.
Fox CW, Bush ML, Wallin WG (2003). Maternal age affects offspring lifespan of the seed beetle, *Callosobruchus maculatus*. *Funct Ecol* **17**: 811–820.
Garcia de Leaniz CG, Fleming IA, Einum S, Verspoor E, Jordan WC, Consuegra S *et al.* (2007). A critical review of adaptive genetic variation in Atlantic salmon: implications for conservation. *Biol Rev* **82**: 173–211.
Keller LF, Waller DM (2002). Inbreeding effects in wild populations. *Trends Ecol Evol* **17**: 230–241.
Koskinen MT, Haugen TO, Primmer CR (2002). Contemporary fisherian life history evolution in small salmonid populations. *Nature* **419**: 826–830.
Kruuk LEB, Slate J, Wilson AJ (2008). New answers for old questions: the evolutionary quantitative genetics of wild animal populations. *Annu Rev Ecol Evol Syst* **39**: 525–548.
Lindholm AK, Hunt J, Brooks R (2006). Where do all the maternal effects go? Variation in offspring body size through ontogeny in the live-bearing fish *Poecilia parae*. *Biol Lett* **2**: 586–589.
Lynch CB (1994). Evolutionary inferences from genetic analysis of cold adaptation in laboratory and wild populations of the house mouse, *Mus domesticus*. In: Boake CRB (ed) *Quantitative Genetic Studies of Behavioral Evolution*. University of Chicago Press: Chicago, IL, USA.
Lynch M, Walsh B (1998). *Genetics and Analysis of Quantitative Traits*. Sinauer Associates: Sunderland, MA, USA.
Metcalfe NB, Thorpe JE (1992). Early predictors of life history events: the link between first feeding date, dominance and seaward migration in Atlantic salmon, *Salmo salar*. *J Fish Biol* **41**: 93–99.
Mousseau TA, Fox CW (1998). The adaptive significance of maternal effects. *Trends Ecol Evol* **13**: 403–407.
Neff BD, Fraser BA (2010). A program to compare genetic differentiation statistics across loci using resampling of individuals and loci. *Mol Ecol Resour* **10**: 546–550.
Neff BD, Pitcher TE (2008). Mate choice for non-additive genetic benefits: a resolution to the lek paradox. *J Theor Biol* **254**: 147–155.
Neff BD, Garner SR, Pitcher TE (2011). Conservation and enhancement of wild fish populations: preserving genetic fitness versus genetic diversity. *Can J Fish Aquat Sci* **68**: 1149–1154.
Pakkasmaa S, Peuhkuri N, Laurila A, Hirvonen H, Ranta E (2001). Female and male contribution to egg size in salmonids. *Evol Ecol* **15**: 143–153.
Pitcher TE, Neff BD (2006). MHC class IIB alleles contribute to both additive and nonadditive genetic effects on survival in *Chinook salmon*. *Mol Ecol* **15**: 2357–2365.
Puurtinen M, Ketola T, Kotiaho JS (2009). The good-genes and compatible-genes benefits of mate choice. *Am Nat* **174**: 741–752.
Qvarnström A, Price TD (2001). Maternal effects, paternal effects and sexual selection. *Trends Ecol Evol* **16**: 95–100.
Räsänen K, Kruuk LEB (2007). Maternal effects and evolution at ecological time-scales. *Funct Ecol* **21**: 408–421.

Roff DA, Emerson K (2006). Epistasis and dominance: evidence for differential effects in life-history versus morphological traits. *Evolution* **60**: 1981–1990.
Schluter D, Clifford EA, Nemethy M, McKinnon JS (2004). Parallel evolution and inheritance of quantitative traits. *Am Nat* **163**: 809–822.
Schumer M, Krishnakant K, Renn SCP (2011). Comparative gene expression profiles for highly similar aggressive phenotypes in male and female cichlid fishes (*Julidochromis*). *J Exp Biol* **214**: 3269–3278.
Turner TL, Bourne EC, Von Wettberg EJ, Hu TT, Nuzhdin SV (2010). Population resequencing reveals local adaptation of *Arabidopsis lyrata* to serpentine soils. *Nat Genet* **42**: 260–263.
Willis JH, Orr HA (1993). Increased heritable variation following population bottlenecks: the role of dominance. *Evolution* **47**: 949–957.
Wilson AJ, Réale D (2006). Ontogeny of additive and maternal genetic effects: lessons from domestic mammals. *Am Nat* **167**: E23–E38.

Supplementary Information accompanies this paper on Heredity website (http://www.nature.com/hdy)

Supplementary Information for: “Genetic architecture of survival and fitness-related traits in two populations of Atlantic salmon”

Appendix A. Details of the broodstock, rearing for families, and methodology used to estimate parameters for two populations of Atlantic salmon (*Salmo salar*).

Appendix B. Summary of survival and fitness-related traits for families from two populations of Atlantic salmon (*Salmo salar*).

Appendix C. Summary of mixed-effects models for the architecture of survival and fitness-related traits in two populations of Atlantic salmon (*Salmo salar*).

Appendix A. Details of the broodstock, rearing for families, and methodology used to estimate parameters for two populations of Atlantic salmon (*Salmo salar*).

Adult broodstock fish from each source population were provided by the Ontario Ministry of Natural Resources (OMNR). Fertilized eggs from single-pair matings of wild LaHave (44°14′N 64°20′W) were received by the OMNR Normandale Fish Culture Station, Vittoria, Ontario from 1989 to 1995 (OMNR, 2005). Captive LaHave generations were produced every year from single-pair matings starting in 1996 at Normandale and then at the OMNR Harwood Fish Culture Station, Harwood, Ontario starting in 2006. Fertilized eggs from single-pair matings of wild Sebago in Panther River (43°53'N, 70°27'W) were received by Harwood in 2006. The populations were raised to maturity at Harwood.

For our study, the adult fish were lightly anaesthetized using tricaine methanesulphonate (MS-222), measured (nearest 0.5 cm) and weighed (nearest 1 g), and then crosses were performed following the methods of Pitcher and Neff (2006). The eggs were then rinsed and disinfected with 0.5% Ovadine for 30 minutes. The disinfection is a government policy in Ontario to minimize the potential establishment or spread of disease within hatcheries, but does not affect egg survival (Fowler and Banks, 1991). Eggs were transported the same day to the OMNR Codrington Research Facility, Codrington, Ontario where they were disinfected again with 0.5% Ovadine for 10 minutes (the second disinfection also does not affect egg survival; Jodun and Millard, 2001). Eggs were supplied with water from a natural spring-fed stream (Marsh Creek) at ambient temperatures for Southern Ontario streams and held in the dark. Dead and unfertilized eggs were counted and removed every two days. Removed eggs from the first month were frozen at -80°C and later soaked in 5% acetic acid for 10 minutes to measure

fertilization success (see Hoysak and Riley, 2001). Only fertilized eggs were used in the measurements of egg survival.

Alevins (555 degree-days from fertilization) were removed from the incubators, counted, and transferred to tanks at an average density of 155 alevins on 1-3 February. Alevins from replicate sections for individual families were pooled from the incubator sections before being transferred to rearing tanks. Tanks contained inserts that divided the area into two sections (each 25 cm × 38 cm) at a water depth of 20 cm (19 L). Individual families were distributed among two to three tank sections based on alevin numbers. Full-sibling replicates were always placed in different tanks. Poultry Astroturf ® substrate (GrassWorx, St. Louis, Missouri), which mimics natural alevin substrate (Fuss and Johnson, 1988), was placed at the bottom of the tanks and held down with stainless steel bars. Alevins continued to be supplied with water from Marsh Creek and the tanks were covered with opaque lids. The Astroturf was removed on 7 March when the alevins had little yolk sac remaining. On 19-22 March, opaque lids were replaced with clear lids on the tanks and all alevins (now called fry, 820 degree-days) were supplied commercial feed (EWOS, Surrey, British Columbia) four times a day *ad libitum*. Dead alevins were counted and removed every two days while in the incubators and every week while in the tanks. Dead fry were similarly counted and removed every week until 15 May when fry (1275 degree-days) were released into the wild.

Egg contents analysis followed the methods of Einum and Fleming (2000). Carbohydrates are low in salmonid eggs (< 0.6%, Kamler, 1992) and were not included in the analysis. Briefly, fat content was determined in seven eggs from each female by the difference between egg dry mass and egg mass after fat extraction. Protein content was next determined by the difference between egg fat-free mass and egg mass after combustion (the ashes). Combustion

was performed at 550°C for 12h (also see Jobling, 1995). Measures of fat, protein, and energy content relative to the wet mass of the egg (i.e., egg value / wet mass of egg) were used for subsequent analyses.

As the eggs hatched, the number of alevins was recorded daily at the same time of day. For replicates that contained large numbers of alevins, digital photographs were taken to determine precise daily numbers of alevins, using ImageJ version 1.38 (NIH, Bethesda, MD, available at www.rsbweb.nih.gov/ij/). Development time to hatch was measured in growing degree-days ($\Delta D = \sum$ ˚C per day) from the spawning date.

At hatch, digital photographs were taken and used to determine alevin body length (length from the tip of the snout to the last trace of visible tail) and yolk sac dimensions (length and width). Ten randomly chosen alevins from each replicate were placed in a Petri dish and ImageJ was calibrated to a ruler that was placed underneath the Petri dish for size measurements (nearest 0.1 mm). Yolk sac volume was calculated as $yolk\ sac\ length \times yolk\ sac\ width^2 \times \pi/6$ (Koskinen *et al.*, 2002). At yolk sac absorption, digital photographs were again taken for body length measurements based 20 randomly chosen fry from each replicate that were placed into two Petri dishes that each contained 10 fry. Specific growth rate (SGR) was calculated as $100 \times [\ln(body\ length\ at\ yolk\ sac\ absorption) - \ln(body\ length\ at\ hatch)]/ \sum$ ˚C per day (Koskinen *et al.*, 2002) and yolk sac conversion efficiency (YCE) was calculated as (*body length at yolk sac absorption - body length at hatch*)/ *yolk sac volume* (Fraser *et al.*, 2010).

Literature Cited

Einum S, Fleming IA (2000). Selection against late emergence and small offspring in Atlantic salmon (*Salmo salar*). *Evolution* **54**: 628-639.

Fowler LG, Banks JL (1991). A safe level of iodophor for treating eggs of fall chinook salmon during water hardening. *Prog Fish-Cult* **53**: 250-251.

Fraser DJ, Houde ALS, Debes PV, O'Reilly P, Eddington JE, Hutchings JA (2010). Consequences of farmed–wild hybridization across divergent wild populations and multiple traits in salmon. *Ecol Appl* **20**: 935-953.

Fuss HJ, Johnson C (1988). Effects of artificial substrate and covering on growth and survival of hatchery-reared coho salmon. *Prog Fish-Cult* **50**: 232-237.

Hoysak DJ, Liley NR (2001). Fertilization dynamics in sockeye salmon and a comparison of sperm from alternative male phenotypes. *J Fish Biol* **58**: 1286-1300.

Jobling M (1995). *Environmental biology of fishes*. Chapman and Hall: London.

Jodun WA, Millard MJ (2001). Effect of iodophor concentration and duration of exposure during water hardening on survival of Atlantic salmon eggs. *N Amer J Aquacult* **63**: 229-233.

Kamler E (1992). Characteristics of fish reproductive products. In: Kamler E (ed) *Early life history of fish: An energetics approach*, Chapman and Hall: London.

Koskinen MT, Haugen TO, Primmer CR (2002). Contemporary fisherian life history evolution in small salmonid populations. *Nature* **419**: 826-830.

[OMNR] Ontario Ministry of Natural Resources (2005). *Stocks catalogue*. Fish and Wildlife Branch, Ontario Ministry of Natural Resources, Ontario, Canada.

Pitcher TE, Neff BD (2006). MHC class IIB alleles contribute to both additive and nonadditive genetic effects on survival in Chinook salmon *Mol Ecol* **15**: 2357-2365.

Appendix B. Summary of survival and fitness-related traits for families from two populations of Atlantic salmon (*Salmo salar*).

Table B1. Summary of egg traits for matriline families from two populations of Atlantic salmon (*Salmo salar*).

Dam	Egg diameter (mm)	Egg mass (g)	Relative fat (g / g of egg)	Relative protein (g / g of egg)	Relative energy (g / g of egg)
LaHave					
1	5.48 ± 0.35	0.1004 ± 0.0106	0.0062 ± 0.0107	0.3485 ± 0.0270	0.3485 ± 0.4483
2	5.89 ± 0.17	0.1080 ± 0.0071	0 ± 0	0.3544 ± 0.0163	0.3544 ± 0.3905
3	5.61 ± 0.27	0.0938 ± 0.0115	0.0024 ± 0.0065	0.3753 ± 0.0289	0.3753 ± 0.8489
4	5.96 ± 0.47	0.1185 ± 0.0122	0.0067 ± 0.0113	0.3849 ± 0.0282	0.3849 ± 0.6018
5	5.67 ± 0.21	0.1047 ± 0.0129	0 ± 0	0.3878 ± 0.0415	0.3878 ± 0.9967
Sebago					
6	5.09 ± 0.38	0.0715 ± 0.0143	0.0082 ± 0.0140	0.3977 ± 0.0464	0.3977 ± 1.0620
7	5.23 ± 0.23	0.0782 ± 0.0099	0.0067 ± 0.0149	0.3722 ± 0.0363	0.3722 ± 0.8545
8	5.21 ± 0.20	0.0891 ± 0.0067	0.0075 ± 0.0129	0.3575 ± 0.038	0.3575 ± 0.6411
9	5.78 ± 0.39	0.1067 ± 0.0100	0.0133 ± 0.0167	0.3847 ± 0.0278	0.3847 ± 0.6710

Presented are means ± 1SD. Sample size was 7 per dam.

Table B2. Summary of survival for families from two populations of Atlantic salmon (*Salmo salar*).

Dam	Sire	Egg survival (%) Day 0-83	Alevin survival (%) Day 84-138	Fry survival (%) Day 139-192	Overall survival (%)
LaHave					
1	1	86.2 ± 2.0	89.5 ± 4.9	44.3 ± 11.6	31.8
1	2	89.6 ± 1.6	79.0 ± 11.7	35.3 ± 17.7	27.1
1	3	85.4 ± 1.7	85.5 ± 2.4	50.2 ± 26.0	40.9
1	4	82.8 ± 1.1	83.1 ± 10.7	66.0 ± 29.7	47.5
1	5	85.4 ± 2.7	86.3 ± 19.9	46.0 ± 2.7	34.3
2	1	62.4 ± 2.9	85.1 ± 7.3	64.2 ± 13.0	33.5
2	2	80.6 ± 5.0	85.9 ± 1.4	65.1 ± 14.0	42.6
2	3	67.5 ± 2.2	84.4 ± 2.7	55.1 ± 10.3	29.4
2	4	61.1 ± 4.2	91.3 ± 6.2	70.1 ± 11.3	40.2
2	5	74.2 ± 7.4	84.9 ± 4.6	30.8 ± 14.7	19.1
3	1	72.9 ± 1.0	88.9 ± 0.2	61.9 ± 9.9	37.7
3	2	75.9 ± 3.8	90.4 ± 5.6	53.2 ± 11.8	36.8
3	3	87.1 ± 2.1	89.8 ± 6.6	61.2 ± 18.0	45.7
3	4	82.8 ± 3.4	90.0 ± 1.4	57.6 ± 13.4	40.2
3	5	79.8 ± 3.4	93.2 ± 1.6	63.2 ± 16.0	42.4
4	1	52.2 ± 4.0	80.0 ± 1.0	85.8 ± 5.4	35.8
4	2	43.7 ± 10.3	67.8 ± 12.3	81.4 ± 6.4	22.7
4	3	36.3 ± 25.5	81.2 ± 2.9	67.5 ± 3.7	18.0
4	4	21.0 ± 2.1	69.4 ± 9.4	91.8 ± 7.8	13.0
4	5	44.0 ± 9.4	83.5 ± 5.7	86.5 ± 0.9	29.8
5	1	73.3 ± 3.8	84.7 ± 4.9	86.2 ± 0.5	52.1
5	2	76.5 ± 2.2	74.7 ± 2.3	76.6 ± 20.1	44.2
5	3	80.5 ± 0.3	82.2 ± 3.6	74.3 ± 9.4	48.7
5	4	73.0 ± 1.3	90.2 ± 2.8	72.2 ± 4.0	46.4
5	5	59.6 ± 12.5	78.4 ± 10.2	69.6 ± 15.6	31.5
Sebago					
6	6	62.5 ± 0.9	79.4 ± 4.5	41.5 ± 9.9	17.1
6	7	66.8 ± 9.9	84.8 ± 2.7	35.5 ± 19.7	20.2
6	8	63.8 ± 3.9	75.3 ± 4.3	54.0 ± 21.1	24.9
6	9	57.5 ± 7.1	75.7 ± 16.4	72.1 ± 4.4	30.5
6	10	63.2 ± 7.3	72.9 ± 2.9	64.7 ± 1.2	29.6
7	6	61.8 ± 24.7	76.3 ± 5.5	34.8 ± 13.6	12.2
7	7	59.7 ± 7.6	85.9 ± 0.2	70.8 ± 39.8	32.5
7	8	58.7 ± 8	78.6 ± 0.7	63.8 ± 13.7	28.5
7	9	70.1 ± 3.4	82.6 ± 8.7	59.6 ± 4.1	33.1

7	10	55 ± 1	79.6 ± 2.7	68.1 ± 1.5	28.9
8	6	41.9 ± 9.8	81.3 ± 0	32.7 ± 0	8.1
8	7	28.8 ± 1.7	69.4 ± 0	62.7 ± 0	11.1
8	8	25.6 ± 3.5	56.8 ± 0	26.1 ± 0	2.6
8	9	16.1 ± 1.2	94.0 ± 0	51.1 ± 0	6.5
8	10	15.8 ± 6.1	62.3 ± 0	39.4 ± 0	2.9
9	6	48.0 ± 29.5	88.8 ± 1.5	45.3 ± 18.2	17.2
9	7	66.1 ± 2.9	72.0 ± 2.0	61.0 ± 1.7	27.1
9	8	75.9 ± 0.8	88.9 ± 1.2	81.5 ± 20.0	52.6
9	9	74.5 ± 10.8	87.5 ± 0.2	79.3 ± 12.9	48.3
9	10	63.1 ± 19.8	88.4 ± 0.4	70.3 ± 9.8	37.8

Presented are means ± 1SD. Sample sizes were the number of replicates: 3 for each LaHave family and 2 for each Sebago family.

Table B3. Summary of fitness-related traits for families from two populations of Atlantic salmon (*Salmo salar*).

Dam	Sire	Development time to hatch (degree-days)	Body length at hatch (mm)	Yolk sac volume (mm^3)	Body length at yolk sac absorption (mm)	Specific growth rate (100 × ln(mm) / degree-days)	Yolk sac conversion efficiency (mm / mm^3)
LaHave							
1	1	480.5 ± 5.2	16.0 ± 0.6	58.56 ± 8.34	26.0 ± 0.9	0.152 ± 0.005	0.171 ± 0.008
1	2	481.9 ± 5.4	16.5 ± 0.5	61.77 ± 9.08	25.2 ± 0.9	0.133 ± 0.007	0.141 ± 0.010
1	3	480.6 ± 4.2	16.0 ± 0.6	63.56 ± 9.15	25.6 ± 0.6	0.148 ± 0.001	0.152 ± 0.001
1	4	481.1 ± 6.1	16.2 ± 0.6	59.44 ± 8.07	25.2 ± 0.7	0.138 ± 0.003	0.152 ± 0.004
1	5	481.6 ± 5.1	16.1 ± 0.6	60.71 ± 10.58	25.5 ± 0.9	0.143 ± 0	0.154 ± 0.001
2	1	480.9 ± 7.2	16.2 ± 0.6	78.32 ± 14.82	26.5 ± 0.8	0.153 ± 0.002	0.131 ± 0.003
2	2	480.3 ± 5.7	16.4 ± 0.7	73.15 ± 12.45	26.1 ± 0.9	0.144 ± 0	0.132 ± 0
2	3	480.7 ± 6.8	16.1 ± 0.6	73.95 ± 17.00	26.2 ± 0.7	0.152 ± 0.002	0.137 ± 0.003
2	4	481.1 ± 6.6	16.3 ± 0.8	74.90 ± 11.10	25.8 ± 0.9	0.143 ± 0.001	0.126 ± 0.001
2	5	480.0 ± 8.4	16.0 ± 0.7	68.44 ± 13.93	26.1 ± 0.8	0.153 ± 0.001	0.148 ± 0.001
3	1	481.9 ± 4.2	16.6 ± 0.4	68.24 ± 14.09	26.3 ± 1.1	0.144 ± 0.013	0.143 ± 0.017
3	2	481.6 ± 4.7	16.5 ± 0.6	64.60 ± 13.90	25.8 ± 0.7	0.140 ± 0	0.145 ± 0
3	3	478.8 ± 4.4	16.4 ± 0.6	68.97 ± 16.18	25.3 ± 0.8	0.136 ± 0.004	0.129 ± 0.005
3	4	477.9 ± 4.7	16.8 ± 0.6	67.45 ± 13.79	25.6 ± 0.9	0.132 ± 0.001	0.131 ± 0.001
3	5	480.0 ± 4.3	16.1 ± 0.6	64.59 ± 11.94	25.4 ± 1.0	0.141 ± 0.004	0.143 ± 0.005
4	1	477.5 ± 9.9	16.8 ± 0.6	86.92 ± 15.87	26.7 ± 0.7	0.145 ± 0	0.114 ± 0
4	2	476.9 ± 9.4	16.7 ± 0.7	83.32 ± 13.81	26.8 ± 0.8	0.149 ± 0	0.122 ± 0
4	3	477.2 ± 7.3	16.3 ± 0.7	95.77 ± 24.34	26.4 ± 0.9	0.151 ± 0.001	0.106 ± 0.001
4	4	476.5 ± 11.0	16.6 ± 0.9	87.16 ± 12.66	26.7 ± 1.0	0.148 ± 0.003	0.116 ± 0.003
4	5	479.2 ± 9.3	16.5 ± 0.9	96.58 ± 15.73	26.6 ± 1.2	0.149 ± 0.004	0.105 ± 0.004
5	1	479.5 ± 4.9	15.8 ± 0.7	62.46 ± 7.83	25.1 ± 0.9	0.145 ± 0	0.149 ± 0
5	2	478.5 ± 5.7	15.8 ± 0.8	69.62 ± 15.76	25.5 ± 1.1	0.149 ± 0.005	0.139 ± 0.006
5	3	478.2 ± 7.7	15.6 ± 0.9	77.71 ± 21.18	25.4 ± 1.0	0.153 ± 0.003	0.126 ± 0.003
5	4	474.7 ± 8.6	15.7 ± 0.6	66.72 ± 11.38	25.1 ± 0.8	0.146 ± 0.001	0.141 ± 0.001
5	5	478.6 ± 6.6	15.2 ± 1.2	70.03 ± 12.27	25.1 ± 1.0	0.156 ± 0.001	0.141 ± 0.001

Sebago							
6	6	477.6 ± 13.1	15.4 ± 0.6	56.87 ± 11.99	24.8 ± 1.1	0.14 ± 0.004	0.165 ± 0.006
6	7	479.7 ± 12.1	15.5 ± 0.8	56.27 ± 9.98	25.2 ± 0.7	0.144 ± 0.003	0.173 ± 0.005
6	8	478.7 ± 9.4	15.2 ± 0.5	58.52 ± 8.83	24.7 ± 0.9	0.143 ± 0.003	0.162 ± 0.004
6	9	479.6 ± 10.1	15.4 ± 0.7	58.13 ± 12.15	25.1 ± 1.3	0.144 ± 0.006	0.167 ± 0.008
6	10	476.3 ± 13.8	15.1 ± 0.7	61.18 ± 13.22	25.5 ± 0.9	0.154 ± 0.009	0.17 ± 0.013
7	6	471.2 ± 12.5	15.5 ± 0.5	52.99 ± 12.69	24.9 ± 0.8	0.139 ± 0	0.177 ± 0
7	7	473.6 ± 10.6	15.4 ± 1.0	61.22 ± 13.99	25.9 ± 0.8	0.154 ± 0.006	0.173 ± 0.009
7	8	471.1 ± 12.5	15.3 ± 0.6	56.72 ± 8.77	25.7 ± 0.7	0.153 ± 0.003	0.183 ± 0.004
7	9	472.2 ± 10.6	15.7 ± 0.6	70.08 ± 13.65	25.3 ± 0.8	0.141 ± 0.007	0.138 ± 0.009
7	10	469.8 ± 11.2	15.3 ± 0.9	63.68 ± 10.93	25.2 ± 0.8	0.146 ± 0.003	0.154 ± 0.003
8	6	473.7 ± 14.0	15.5 ± 0.7	58.68 ± 10.8	25.2 ± 0.9	0.144 ± 0.009	0.166 ± 0.013
8	7	471.2 ± 14.9	15.2 ± 0.6	58.82 ± 8.56	25.6 ± 0.5	0.152 ± 0.002	0.176 ± 0.003
8	8	471.4 ± 14.7	15.4 ± 0.7	60.80 ± 11.87	25.5 ± 1.0	0.149 ± 0.007	0.166 ± 0.010
8	9	474.9 ± 13.3	16.8 ± 0.4	70.01 ± 11.16	25.4 ± 0.7	0.122 ± 0.007	0.123 ± 0.008
8	10	462.8 ± 16.4	15.3 ± 0.7	66.49 ± 9.74	25.2 ± 0.8	0.147 ± 0.005	0.150 ± 0.007
9	6	471.1 ± 10.9	15.9 ± 0.5	78.12 ± 18.35	27.1 ± 0.8	0.158 ± 0.007	0.144 ± 0.008
9	7	467.6 ± 12.0	16.3 ± 0.8	82.32 ± 12.78	26.9 ± 0.6	0.147 ± 0.003	0.129 ± 0.003
9	8	470.6 ± 10.5	16.0 ± 0.7	68.44 ± 16.51	26.6 ± 1.0	0.149 ± 0.003	0.155 ± 0.004
9	9	473.6 ± 10.8	16.0 ± 0.6	72.11 ± 11.48	26.9 ± 0.9	0.154 ± 0.005	0.152 ± 0.006
9	10	468.2 ± 10.5	16.1 ± 0.6	81.42 ± 19.21	26.8 ± 0.8	0.151 ± 0.009	0.132 ± 0.010

Presented are means ± 1SD. Development time to hatch, specific growth rate, and yolk sac volume sample sizes were the number of replicates: 3 for each LaHave family and 2 for each Sebago family. Body length at hatch, yolk sac volume at hatch, and body length at yolk sac absorption sample sizes were 10 offspring per replicate: 30 for each LaHave family and 20 for each Sebago family.

Appendix C. Summary of mixed-effects models for the architecture of survival and fitness-related traits in two populations of Atlantic salmon (*Salmo salar*).

Trait	*n*	*p*-value	σ^2 (% total variance)	phenotypic variance	% phenotypic variance
Egg survival (Day 0-83)					
LaHave					
dam	5	< 0.001	0.499 (28.7)	maternal	22.3
sire	5	0.5654	0.015 (0.9)	additive	2.8
dam × sire	25	< 0.001	0.132 (7.6)	non-additive	24.5
tray	13	< 0.001	0.086 (5.0)		
Residual			1.002 (57.7)		
Sebago					
dam	4	< 0.001	0.988 (24.5)	maternal	12.7
sire	5	0.8746	0.039 (1.0)	additive	2.1
dam × sire	20	< 0.001	1.125 (27.9)	non-additive	60.1
tray	13	< 0.001	0.887 (22.0)		
Residual			0.992 (24.6)		
Alevin survival (Day 84-138)					
LaHave					
dam	5	0.036	0.010 (7.7)	maternal	5.3
sire	5	0.719	0.012 (0.9)	additive	3.4
dam × sire	25	0.286	0.089 (7)	non-additive	20.8
tank	38	0.044	0.126 (9.8)		
Residual			0.967 (74.8)		
Sebago					
dam	4	0.524	0.093 (5.9)	maternal	4.7
sire	5	0.692	0.019 (1.3)	additive	4.6
dam × sire	20	0.500	0.076 (6.5)	non-additive	18.5
tank	31	0.084	0.236 (15.5)		
Residual			0.950 (70.9)		
Fry survival (Day 139-192)					
LaHave					
dam	5	0.090	0.173 (8.6)	maternal	5.9
sire	5	1	0 (0.0)	additive	0.0
dam × sire	25	< 0.001	0.308 (15.3)	non-additive	41.9
tank	51	< 0.001	0.550 (27.3)		
Residual			0.984 (48.8)		
Sebago					
dam	4	0.161	0.137 (7.1)	maternal	0
sire	5	0.114	0.235 (12.1)	additive	34.2
dam × sire	20	0.287	0.078 (4.0)	non-additive	11.4
tank	32	< 0.001	0.526 (27.0)		
Residual			0.970 (49.8)		
Development time to hatch					
LaHave					
dam	5	0.003	1.58 (3.8)	maternal	1.9
sire	5	0.046	0.72 (1.7)	additive	6.2
dam × sire	25	< 0.001	1.00 (2.4)	non-additive	8.7
tray	13	< 0.001	1.82 (4.3)		
Residual			36.72 (87.8)		

Sebago					
dam	4	0.004	12.53 (8.0)	maternal	4.9
sire	5	0.057	4.03 (2.6)	additive	9.2
dam × sire	20	0.025	3.42 (2.2)	non-additive	7.8
tray	13	< 0.001	4.22 (2.7)		
Residual			132.49 (84.5)		
Body length at hatch					
LaHave					
dam	5	< 0.001	0.134 (21.6)	maternal	17.1
sire	5	0.038	0.018 (2.9)	additive	10.8
dam × sire	25	0.365	0.006 (1.0)	non-additive	4.0
tray	13	0.121	0.007 (1.1)		
Residual			0.453 (73.3)		
Sebago					
dam	4	0.124	0.078 (12.1)	maternal	7.8
sire	5	0.969	0.005 (0.8)	additive	2.3
dam × sire	20	< 0.001	0.092 (14.4)	non-additive	39.7
tray	13	0.194	0.017 (2.7)		
Residual			0.448 (70.0)		
Yolk sac volume					
LaHave					
dam	5	< 0.001	117.3 (36.4)	maternal	33.3
sire	5	0.536	1.73 (0.5)	additive	2.0
dam × sire	25	0.028	7.41 (2.3)	non-additive	8.5
tray	13	0.728	0.67 (0.2)		
Residual			194.7 (60.5)		
Sebago					
dam	4	0.001	60.6 (25.2)	maternal	19.0
sire	5	0.461	4.83 (2.0)	additive	6.6
dam × sire	20	0.007	14.4 (6.0)	non-additive	19.7
tray		0.604	1.52 (0.6)		
Residual			159.4 (66.2)		
Body length at yolk sac absorption					
LaHave					
dam	5	<0.001	0.273 (24.2)	maternal	20.4
sire	5	0.347	0.015 (1.3)	additive	4.7
dam × sire	25	0.022	0.036 (3.2)	non-additive	11.4
tank	38	<0.001	0.042 (3.8)		
Residual			0.760 (67.5)		
Sebago					
dam	4	< 0.001	0.578 (39.2)	maternal	33.4
sire	5	0.993	0 (0.0)	additive	0.0
dam × sire	20	0.002	0.086 (5.8)	non-additive	19.8
tank	31	< 0.001	1.29 (8.8)		
Residual			0.680 (46.2)		
Specific growth rate					
LaHave					
dam	5	0.022	1.9e-5 (35.1)	maternal	11.5
sire	5	0.195	6.9e-6 (13.0)	additive	26.8
dam × sire	25	0.036	1.2e-5 (22.5)	non-additive	46.6
tank	38	0.943	2.4e-7 (0.5)		
Residual			1.6e-5 (29.3)		

Sebago					
dam	4	1	2.8e-6 (3.6)	maternal	1.3
sire	5	1	0 (0.0)	additive	0.0
dam × sire	20	0.004	4.5e-5 (57.2)	non-additive	84.2
tank	31		0 (0.0)		
Residual			3.1e-5 (39.2)		
Yolk sac conversion efficiency					
LaHave					
dam	5	< 0.001	2.1e-4 (71.9)	maternal	43.9
sire	5	0.564	7.4e-6 (2.5)	additive	6.4
dam × sire	25	0.0002	5.2e-5 (17.7)	non-additive	44.8
tank	38	1	0 (0.0)		
Residual			2.3e-5 (7.9)		
Sebago					
dam	4	0.274	9.0e-5 (25.7)	maternal	6.0
sire	5	0.618	3.4e-5 (9.7)	additive	14.5
dam × sire	20	0.0001	1.7e-4 (48.9)	non-additive	73.5
tank	31	1	0 (0.0)		
Residual			5.5e-5 (15.8)		

Presented are the results on the observed data for both populations. All mixed-effects models contained a random effect for position effects (i.e., *tray ID* or *tank ID*).